# In Vitro Systems for the Assessment of Chronic Cardiotoxic Effects: News from the HESI Stem Cell Working Group

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#### Abstract

Since long-term exposure of cancer-related therapeutics have been linked to alterations of cardiac function in patients, the Stem Cell Working Group as part of the Health and Environmental Science Institute (HESI) endeavors to gain further insight into chronic cardiotoxicity. The objective of the study was to optimize non-clinical safety assessment strategies of chronic cardiotoxicity by testing prolonged exposure of reference compounds on cell-based assay systems using human induced pluripotent stem cell-derived cardiomyocytes (hiPSC-CMs).



## **Study Outline**

A panel of 12 compounds was selected to evaluate chronic adverse side effects on hiPSC-CMs, including alterations in energetics and contraction force as well as electrophysiological and structural disturbance.

For the quantification of adverse side effects, a variety of methods was used, covering electrophysiology, mechanobiology, optical methods as well as biochemical assays.

<b>Target Toxicity</b>	Compound	Conc. Range	Parameter	Method
Energetics/ Mitochondrial Toxicity	Doxorubcin	0.1 - 3 μΜ	Electrophysiology	Multielectrode array
	Erlotinib	0.3 - 10 μM	Liectrophysiology	Impedance
	Sunitib	0.01 - 1 μM	Mechanobiology	Contraction force
Electrophysiological Disturbance	Pentamidine	0.1 - 3 μΜ		Optical contraction analysis
	Arsenic Trioxide	0.1 - 3 μΜ	Optical Methods	Voltage sensitive dye
Contractility	BMS-986094	0.1 - 3 μΜ		Calcium imaging
	Milrinone	0.1 - 10 μΜ	Biochemical	Oxygen consumption rate
	Nioltinib	0.1 - 10 μM	Biochennical	Biomarkers
Structural/myofilament Disturbance	Endothelin-1	0.3 - 100 nM		green: presented in this poste
	Vinblastine	1 - 300 nM	<b>个 Table 2:</b> Methods	used for the quantification of chronic
	Vincristine	1 - 300 nM	cardiotoxic compound effects	
	Vinorelbine	0.1 - 3 μM	← Table 1: Cardiotoxic compounds sorted by target	
	green: p	resented in this poster	toxicity	

## Technology

**iCell Cardiomyocytes**<sup>2</sup> are highly pure human iPSC-derived cardiomyocytes and allow for robust and reproducible quantification of compound effects as demonstrated in the CiPA study (Blinova *et al.*, 2017).

The **CardioExcyte 96** is an impedance-based system where hiPSC-derived cardiomyocytes are cultured on gold electrodes, enabling label-free and continuous quantification of electrical impedance as a measure of monolayer integrity.

In the **FLEXcyte 96** system, hiPSC-derived cardiomyocytes are cultured on ultra-thin silicone membranes, mimicking the mechanical environment of native human heart tissue. Rhythmic contractions deflect the membranes, quantified by capacitive distance sensing. Parameters analyzed are contractile force (AMP), beat rate (BR), beat rate regularity (BRR), rising time (RT), falling time (FT). (Gossmann *et al.*, 2016 and 2020).

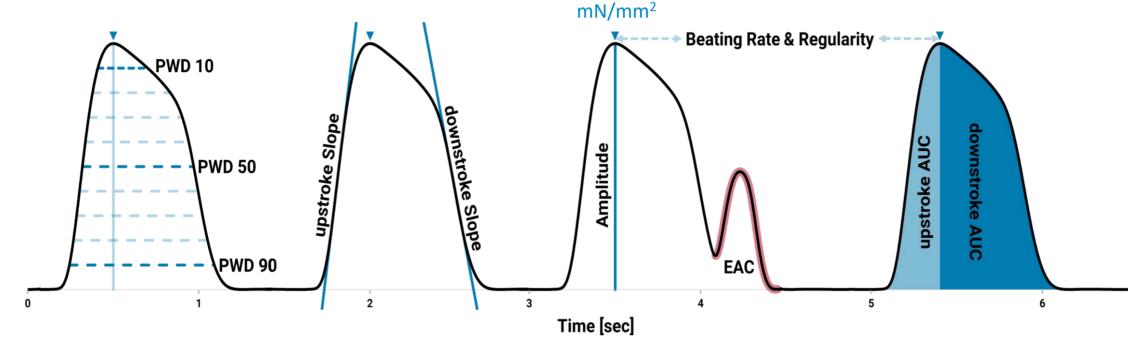
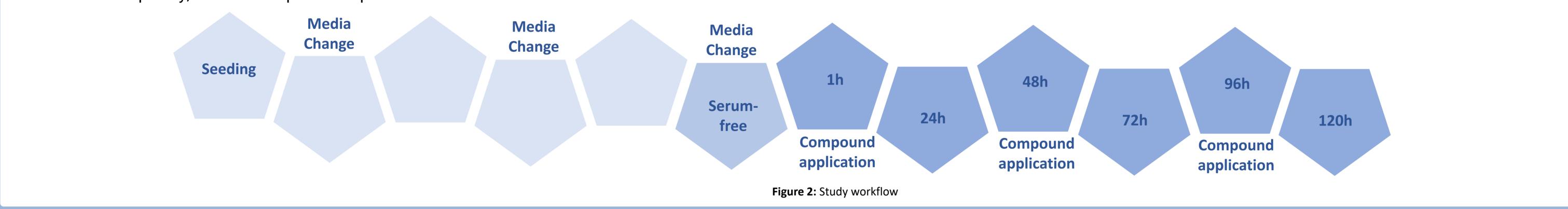


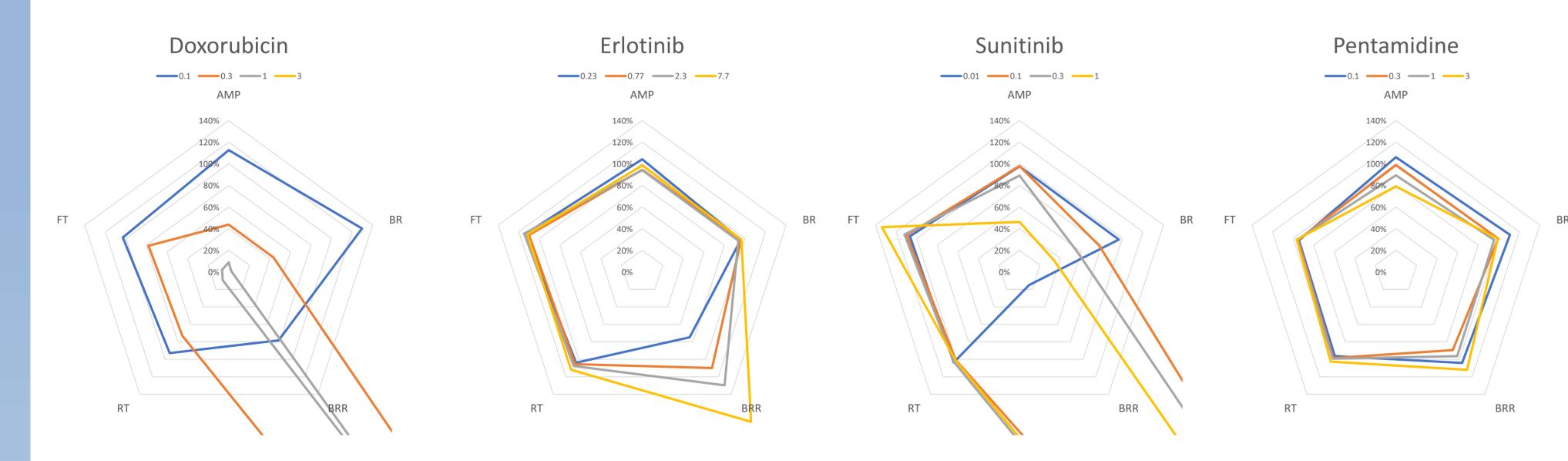
Figure 1: FLEXcyte 96 contraction analysis parameters

# Workflow

hiPSC-CMs were seeded from cryopreserved state into the assay format and cultured in maintenance medium for 5 days. 24h prior to compound application, medium was changed to serum-free, protein-containing assay medium. Baseline measurements were performed before compound application. Compounds were added 4x concentrated during a partial media change. The first compound measurement was carried out after 1h. Subsequently, chronic compound responses were recorded in 24h intervals for a total of 120h.



#### Results



<b>Target Toxicity</b>	Compound	<b>C1</b>	<b>C2</b>	<b>C3</b>	<b>C4</b>
Energetics/	Doxorubcin	0,75	0,45	0,40	0,36
Mitochondrial Toxicity	Erlotinib Sunitib	0,95	0,99	1,03	1,00
Ephys Disturbance	Pentamidine	0,94 0,94	1,10 0,94	1,23 0,81	1,33 0,62
Contractility	BMS-986094	0,93	0,78	0,83	0,77
,	Nilotinib	0,96	1,09	1,14	1,00
Structural/myofilament Disturbance	Endothelin-1	0,82	0,83	0,86	0,87
Disturbance	Vincristine	0,90	0,57	0,43	0,46

Table 3: Effect of the tool compounds on the base impedance as a measure of monolayer integrity

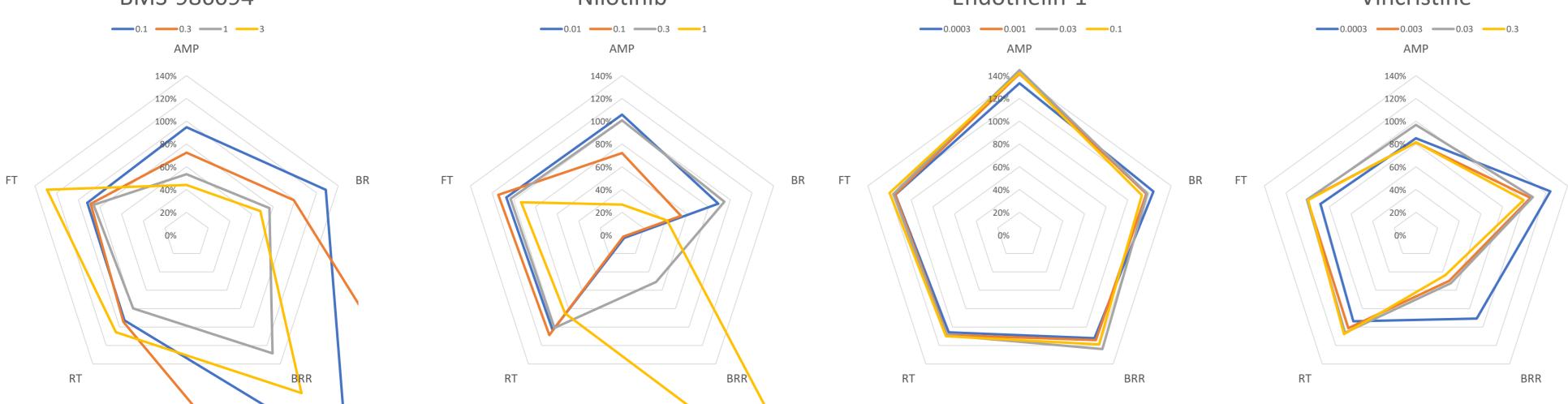
• **Doxorubicin** had a strong toxic effect, measured both in contraction force as well as monolayer integrity.

BMS-986094

Nilotinib

Endothelin-1

Vincristine



- Figure 3: Contraction parameter analysis at 72h post compound application. Parameters: Contraction force (AMP), Beat rate (BR), Beat rate regularity (BRR), Rising time (RT), Falling time (FT)
- Erlotinib had only slight effects on the BRR of hiPSC-CMs and no effect on monolayer integrity.
- Sunitinib had a strong effect on the chronotropy (BR, BRR), but rather low effect on the contractile force. The monolayer integrity was increased.
- Pentamidine had a strong effect on the contraction force and on the monolayer integrity.
- BMS-986094 had a major impact on all contractile parameters and on the monolayer integrity.
- Nilotinib had a strong effect on the contractile parameters, while the monolayer integrity remained unchanged.
- Endothelin-1 had a positive effect on the contraction force, but little or no effect on the monolayer integrity.
- Vincristine had minor effects on the contractile parameters, but a major effect on the monolayer integrity.

#### **References:**

- Blinova, K., Stohlman, J., Vicente, J., Chan, D., Johannesen, L., Hortigon-Vinagre, M. P., Zamora, V., Smith, G., Crumb, W. J., Pang, L., Lyn-Cook, B., Ross, J., Brock, M., Chvatal, S., Millard, D., Galeotti, L., Stockbridge, N., & Strauss, D. G. (2017). Comprehensive Translational Assessment of Human-Induced Pluripotent Stem Cell Derived Cardiomyocytes for Evaluating Drug-Induced Arrhythmias. *Toxicological sciences : an official journal of the Society of Toxicology*, 155(1), 234–247.
- Goßmann, M., Frotscher, R., Linder, P., Neumann, S., Bayer, R., Epple, M., Staat, M., Artmann, G. M. (2016). Mechano-Pharmacological Characterization of Cardiomyocytes Derived from Human Induced Pluripotent Stem Cells. Cellular physiology and biochemistry : international journal of experimental cellular physiology, biochemistry, and pharmacology, 38(3), 1182–1198.
- Goßmann, M., Linder, P., Thomas, U., Juhasz, K., Lemme, M., George, M., Fertig, N., Dragicevic, E., & Stoelzle-Feix, S. (2020). Integration of mechanical conditioning into a high throughput contractility assay for cardiac safety assessment. Journal of pharmacological and toxicological methods, 105, 106892.

